



# Genome Sequencing Identified a SARS-CoV-2 Lineage B.1.1.7 Strain with a High Number of Mutations from Dhaka, Bangladesh

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**ABSTRACT** We report a coding-complete genome sequence of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) strain SARS-CoV-2/BGD/GC001, isolated from a Bangladeshi patient with respiratory symptoms. Phylogenetic analysis assigned this strain to lineage B.1.1.7, which presented a total of 36 mutations in the spike and other genomic regions compared to strain Wuhan Hu-1 (GenBank accession number [NC\\_045512.2](https://www.ncbi.nlm.nih.gov/nuccore/NC_045512.2)).

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) belongs to the family *Coronaviridae* and genus *Betacoronavirus* (1). Lineage B.1.1.7, which emerged in the United Kingdom, has attracted particular attention due to its high transmissibility and immune escape potential (2). Herein, we announce a coding-complete genome sequence of the strain SARS-CoV-2/BGD/GC001 (GC001) belonging to lineage B.1.1.7 and detected in a patient with respiratory symptoms who presented on 31 January 2021 in Dhaka, Bangladesh. Ethical approval was obtained from the icddr,b Research and Ethical Review Committee (protocol number PR-21040).

The specimen (nasopharyngeal swabs) from the symptomatic patient was processed for nucleic acid isolation utilizing a QIAamp viral RNA minikit (Qiagen, Germany). SARS-CoV-2 was confirmed using a TaqMan real-time PCR (RT-PCR) assay (3). A cDNA library was prepared utilizing the Illumina TruSeq stranded total RNA Gold low-throughput (LT) library preparation kit following the manufacturer's instructions; rRNA reduction was performed using the Ribo-Zero Gold protocol. The libraries were normalized to 10 nM following Illumina's standard normalization method and pooled using 10  $\mu$ l of each library, which was then sequenced using the NextSeq v2.5 midoutput kit (2  $\times$  150 cycles) on the NextSeq 500 instrument at the Genomics Center of the icddr,b in Dhaka, Bangladesh. There were 112,690,904 paired-end raw reads, whose quality was assessed using FastQC v0.11.11 (4). The adapters were trimmed using Trimmomatic v0.39 based on Q30 values with the following parameters: window size, 4; Phred quality, 15; and minimum read length, 40 (5). After trimming, 205,697,670 reads were used for reference-based alignment. The SARS-CoV-2-specific reads were mapped and filtered using SMALT v0.7.6 (<http://www.sanger.ac.uk/science/tools/smalt-0>) and SAMtools v1.9 (6). *De novo* assembly was performed with 17,324 reads using SPAdes v3.11.1 (7), and the quality was assessed using QUAST v5.0.2 (8). RATT was employed to annotate the genome with Wuhan-Hu-1 as the reference strain (GenBank accession number [NC\\_045512.2](https://www.ncbi.nlm.nih.gov/nuccore/NC_045512.2)) (9). Mutations were assessed utilizing the Genome Detective Coronavirus Typing Tool (10). The Nextstrain and PANGOLIN Web tools were used for comparative genomics (11, 12). MUSCLE v3.8.31 (13) and MEGA7 (14) software were used to generate a phylogenetic tree using the neighbor-joining method with

**Citation** Mazumder R, Abdullah A, Hossain ME, Rahman MM, Bin Manjur OH, Rahman M, Mondal D. 2021. Genome sequencing identified a SARS-CoV-2 lineage B.1.1.7 strain with a high number of mutations from Dhaka, Bangladesh. *Microbiol Resour Announc* 10:e00345-21. <https://doi.org/10.1128/MRA.00345-21>.

**Editor** John J. Dennehy, Queens College CUNY

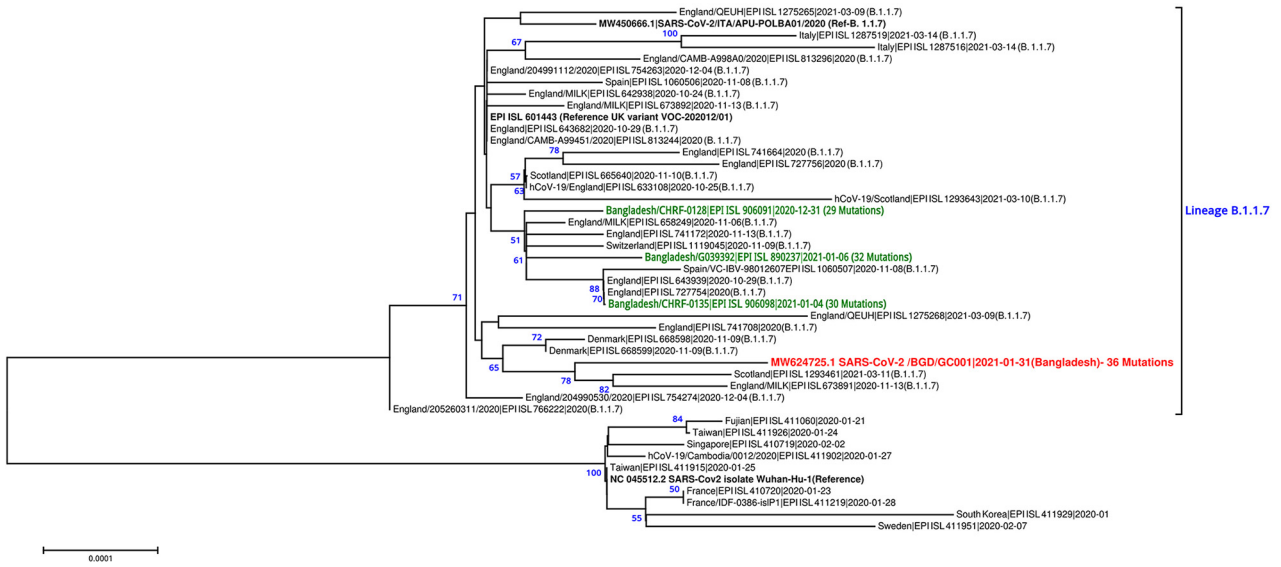
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**Received** 16 April 2021

**Accepted** 27 April 2021

**Published** 27 May 2021



**FIG 1** Phylogenetic tree showing the genome sequence of SARS-CoV-2/BGD/GC001/2021 (GenBank accession number [MW624725.1](#)) together with 44 sequences, including 34 lineage B.1.1.7 sequences and 10 non-B.1.1.7 lineage sequences retrieved from GISAID and NCBI GenBank. The sequence reported in this announcement is highlighted in red; other B.1.1.7 lineage sequences from Bangladesh are shown in green. The percent bootstrap support values are indicated at each node (values of <50 are omitted).

**TABLE 1** Mutations of SARS-CoV-2 strain GC001 (GenBank accession number [MW624725.1](#)) in comparison to the reference strain ([NC\\_045512.2](#))

Gene or region	Mutation no.	CDS codon position <sup>a</sup>	Amino acid change	Nucleotide position	Nucleotide change
5' untranslated region	1			241	C > T
ORF1ab	2	216		913	C > T
	3	615		2110	C > T
	4	924		3037	C > T
	5	1001	T > I	3267	C > T
	6	1708	A > D	5388	C > A
	7	1907		5986	C > T
	8	2230	I > T	6954	T > C
	9	2573		7984	T > C
	10	3198		9857	C > T
	11	3675–3677	SGF deletion	11288–11296	Deletion TCTGGTTTT
	12	4619	P > L	14120	C > T
	13	4715	P > L	14408	C > T
	14	4804		14676	C > T
	15	505		15279	C > T
	16	5304		16176	T > C
	17	6376	P > S	19390	C > T
S	18	69–70	HV deletion	21766–21771	Deletion ACATGT
	19	75	G > V	21786	G > T
	20	144	Y deletion	21992–21994	Deletion TAT
	21	501	N > Y	23063	A > T
	22	570	A > D	23271	C > A
	23	614	D > G	23403	A > G
	24	681	P > H	23604	C > A
	25	716	T > I	23709	C > T
	26	982	S > A	24506	T > G
	27	1118	D > H	24914	G > C
ORF8	28	27	Q > stop	27972	C > T
	29	52	R > I	28048	G > T
	30	68	K > stop	28095	A > T
	31	73	Y > C	28111	A > G
N	32	3	D > L	28280–28282	GAT > CTA
	33	203	R > K	28881–28882	GG > AA
	34	204	G > R	28883	G > C
	35	235	S > F	28977	C > T
	36	269		29080	T > C

<sup>a</sup> CDS, coding DNA sequence.

1,000 bootstraps (Fig. 1). Default parameters were applied for all tools unless otherwise mentioned.

The genome sequence of strain GC001 was 29,842 bp long with a G+C content of 37.98%. Phylogenetic analysis assigned strain GC001 to lineage B.1.1.7, which is a predominant lineage worldwide (Fig. 1). We identified 33 mutations and 3 deletions in GC001 in comparison to strain Wuhan-Hu-1 (GenBank accession number [NC\\_045512.2](https://doi.org/10.1093/cmb.2012.0021)) (Table 1). Strain GC001 showed a high number of mutations compared to other SARS-CoV-2 lineage B.1.1.7 strains identified in Bangladesh at the time of the analysis (Fig. 1). Moreover, strain GC001 did not cluster closely with other B.1.1.7 genomes identified from Bangladesh to date, which hints toward its independent introduction into the country. These observations suggest the importance of genome sequencing of SARS-CoV-2 samples from travelers, particularly those returning from high-risk countries.

**Data availability.** The genome sequence of SARS-CoV-2/BGD/GC001 was deposited in the NCBI database under the BioProject accession number [PRJNA702998](https://doi.org/10.1093/cmb.2012.0021), BioSample accession number [SAMN17993365](https://doi.org/10.1093/cmb.2012.0021), and GenBank accession number [MW624725.1](https://doi.org/10.1093/cmb.2012.0021). The Illumina raw reads have been deposited in the NCBI Sequence Read Archive under accession number [SRR13744683](https://doi.org/10.1093/cmb.2012.0021).

## ACKNOWLEDGMENTS

This research study was funded by core donors who provide unrestricted support to icddr,b for its operations and research. Current donors providing unrestricted support include the governments of Bangladesh, Canada, Sweden, and the United Kingdom. We gratefully acknowledge our core donors for their support and commitment to icddr,b's research efforts.

## REFERENCES

1. Wu F, Zhao S, Yu B, Chen Y-M, Wang W, Song Z-G, Hu Y, Tao Z-W, Tian J-H, Pei Y-Y, Yuan M-L, Zhang Y-L, Dai F-H, Liu Y, Wang Q-M, Zheng J-J, Xu L, Holmes EC, Zhang Y-Z. 2020. A new coronavirus associated with human respiratory disease in China. *Nature* 579:265–269. <https://doi.org/10.1038/s41586-020-2008-3>.
2. Rondinone V, Pace L, Fasanella A, Manzulli V, Parisi A, Capobianchi MR, Ostuni A, Chironna M, Caprioli E, Labonia M, Cipolletta D, Della Rovere I, Serrecchia L, Petrucci F, Pennuzzi G, Galante D. 2021. VOC 202012/01 variant is effectively neutralized by antibodies produced by patients infected before its diffusion in Italy. *Viruses* 13:276. <https://doi.org/10.3390/v13020276>.
3. Lu X, Wang L, Sakthivel SK, Whitaker B, Murray J, Kamili S, Lynch B, Malapati L, Burke SA, Harcourt J, Tamin A, Thornburg NJ, Villanueva JM, Lindstrom S. 2020. US CDC real-time reverse transcription PCR panel for detection of severe acute respiratory syndrome coronavirus 2. *Emerg Infect Dis* 26:1654–1665. <https://doi.org/10.3201/eid2608.201246>.
4. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
5. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
6. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>.
7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
8. Mikheenko A, Pribelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUAST-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>.
9. Otto TD, Dillon GP, Degraeve WS, Berriman M. 2011. RATT: Rapid Annotation Transfer Tool. *Nucleic Acids Res* 39:e57. <https://doi.org/10.1093/nar/gkq1268>.
10. Cleemput S, Dumon W, Fonseca V, Abdool Karim W, Giovanetti M, Alcantara LC, Deforche K, De Oliveira T. 2020. Genome Detective Coronavirus Typing Tool for rapid identification and characterization of novel coronavirus genomes. *Bioinformatics* 36:3552–3555. <https://doi.org/10.1093/bioinformatics/btaa145>.
11. Hadfield J, Megill C, Bell SM, Huddleston J, Potter B, Callender C, Sagulenko P, Bedford T, Neher RA. 2018. Nextstrain: real-time tracking of pathogen evolution. *Bioinformatics* 34:4121–4123. <https://doi.org/10.1093/bioinformatics/bty407>.
12. Rambaut A, Holmes EC, O'Toole Á, Hill V, McCrone JT, Ruis C, Du Plessis L, Pybus OG. 2021. Addendum: a dynamic nomenclature proposal for SARS-CoV-2 lineages to assist genomic epidemiology. *Nat Microbiol* 6:415. <https://doi.org/10.1038/s41564-021-00872-5>.
13. Edgar RC. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 32:1792–1797. <https://doi.org/10.1093/nar/gkh340>.
14. Kumar S, Stecher G, Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol* 33:1870–1874. <https://doi.org/10.1093/molbev/msw054>.